

## Review

## The dorsal raphe nucleus in the control of energy balance

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**Energy balance is orchestrated by an extended network of highly interconnected nuclei across the central nervous system. While much is known about the hypothalamic circuits regulating energy homeostasis, the ‘extra-hypothalamic’ circuits involved are relatively poorly understood. In this review, we focus on the brainstem’s dorsal raphe nucleus (DRN), integrating decades of research linking this structure to the physiologic and behavioral responses that maintain proper energy stores. DRN neurons sense and respond to interoceptive and exteroceptive cues related to energy imbalance and in turn induce appropriate alterations in energy intake and expenditure. The DRN is also molecularly differentiable, with different populations playing distinct and often opposing roles in controlling energy balance. These populations are integrated into the extended circuit known to regulate energy balance. Overall, this review summarizes the key evidence demonstrating an important role for the DRN in regulating energy balance.**

### From hypothalamic to brainstem circuits regulating energy balance

All organisms require stable energy stores, severe deviations of which are incompatible with life. Animals have evolved numerous ways to meet energy demands: Energy is acquired through feeding and expended through external work (locomotion) or internal heat (thermogenesis). These modalities thus offer the organism behavioral and physiologic means to respond to homeostatic threats to peripheral energy stores. In mammals, neurons in several brain areas are known to coordinate and drive responses to deviations in peripheral metabolism.

Decades of pioneering work has focused on the hypothalamus, uncovering an assortment of cell types known to regulate energy balance, such as agouti-related peptide (AgRP) and pro-opiomelanocortin (POMC) neurons in the arcuate nucleus of the hypothalamus (ARC) [1,2]. However, it has become apparent that circuits outside of the hypothalamus also play a pivotal role in energy homeostasis, including neurons distributed across several brainstem loci. Among these neurons are neurons expressing calcitonin gene-related peptide and prodynorphin in the parabrachial nucleus (PBN) [3,4], along with multiple cell types within the nucleus of the solitary tract (NTS) [5,6].

One nucleus that has received considerably less attention, but likely plays an important role in energy balance, is the dorsal raphe nucleus (DRN). The DRN is a midline structure in the dorsal midbrain that borders the ventrolateral periaqueductal gray (VIPAG) and sits immediately beneath the cerebral aqueduct. Subsets of DRN neurons have been implicated in diverse behavioral and physiologic processes, including those related to sociability, anxiety, depression, nociception, and sleep [7–12]. Only recently, however, has this structure received significant emphasis within the setting of energy balance.

Although it is one of several distinct serotonergic raphe nuclei in the mammalian brainstem, the DRN contains the largest group of serotonin (5-HT) neurons in the brain [13,14]. In addition to

### Highlights

The dorsal raphe nucleus (DRN) has extensive afferent and efferent connections with forebrain loci regulating feeding and thermogenesis; the DRN also polysynaptically innervates brown adipose tissue (BAT).

Modulating these DRN projections can augment or suppress food intake and/or thermogenesis.

DRN subpopulations expressing specific neuropeptides and/or their receptors have been functionally implicated in energy balance.

Reduced serotonin (5-HT) release from the DRN increases food intake, and 5-HT release from the DRN is necessary for maintaining normal thermogenesis.

Excitatory and inhibitory DRN neurons respond to energy deficit or surfeit and exert bidirectional control over energy balance.

Inhibitory DRN neurons are responsive to thermal challenge; these neurons regulate BAT thermogenesis and overall energy expenditure.

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5-HT neurons, other principal DRN cell types include GABAergic, dopaminergic (DA) [15–17], and glutamatergic neurons that express the vesicular glutamate transporter (VGLUT) isoforms VGLUT2 and VGLUT3 [11,15,18,19].

While the DRN provides a large proportion of serotonergic innervation to the forebrain, much of the research on central 5-HT in energy balance – with notable exceptions [5] – has focused on post-synaptic targets for 5-HT in the hypothalamus [20]. Other studies on central 5-HT in energy homeostasis, at least historically, have relied upon relatively nonspecific manipulations, such as systemic or intracerebroventricular drug administration. Nonetheless, evidence has steadily emerged supporting a role for the DRN, in and of itself, as a critical locus in energy homeostasis, encompassing both serotonergic and nonserotonergic DRN cell types.

In this review, we summarize the literature which, in aggregate, indicates that DRN neurons play a vital role in maintaining energy balance. This work demonstrates that DRN neurons can sense and respond to physiologically relevant cues, such as hunger or ambient warmth; that subsets of DRN neurons play distinct roles in controlling food intake and energy expenditure; and that these neurons are embedded within broader circuits known to regulate energy balance. Elucidating the DRN's role in regulating energy balance will require further exploration of its molecular, cellular, and circuit-level properties. Characterizing how DRN subpopulations integrate and encode energy-related signals to drive appropriate behavioral or physiologic responses will lead to a better understanding of the neural mechanisms regulating energy balance.

### The role of the DRN in energy intake

Early electrolytic lesion studies of the raphe suggested that the mammalian DRN could play a role in food intake [21]. Consistent with these findings, acutely and nonspecifically inhibiting DRN neurons via the infusion of the GABA<sub>A</sub> receptor agonist muscimol has long been reported to increase feeding in sated rodents [22–24]. Over time, additional neuroanatomical and functional evidence has established a critical role for both serotonergic and nonserotonergic DRN neurons in regulating food intake (Table 1). These cell types also play important roles in motivated behavior, which could relate to both the drive to consume food and the hedonic aspects of feeding (Box 1).

Neuroanatomical tracing studies have shown that the DRN has both extensive afferent and efferent connections with loci known to regulate feeding. With respect to afferent connectivity, the DRN receives significant input from the hypothalamus and extended amygdala, including the central amygdala (CeA), bed nucleus of the stria terminalis (BNST), and the lateral hypothalamus (LH) [25–28]. Notably, activating inhibitory projections from the LH and BNST to the DRN and neighboring vPAG can promote feeding (Figure 1) [24].

The DRN also receives input from AgRP-expressing ‘hunger’ neurons in the ARC. AgRP neurons projecting to the DRN/vPAG are activated by ghrelin and food deprivation, and activation of

Table 1. Alterations in energy balance are driven by signaling from broad dorsal raphe nucleus cell types

	Serotonergic, DRN <sup>5-HT</sup>	Glutamatergic, DRN <sup>VGLUT3</sup>	GABAergic, DRN <sup>GABA</sup>
Food intake	Decrease [23,53–57,66]	Decrease [23]	Increase [23,77,78] Decrease [24]
Energy expenditure	Increase [82,92]	Not known	Decrease [77,78]
Locomotion	Increase [103,104,106] Decrease [105–107]	Increase [23]	Increase [106] Decrease [23,77,106]

Abbreviation: DRN, dorsal raphe nucleus.

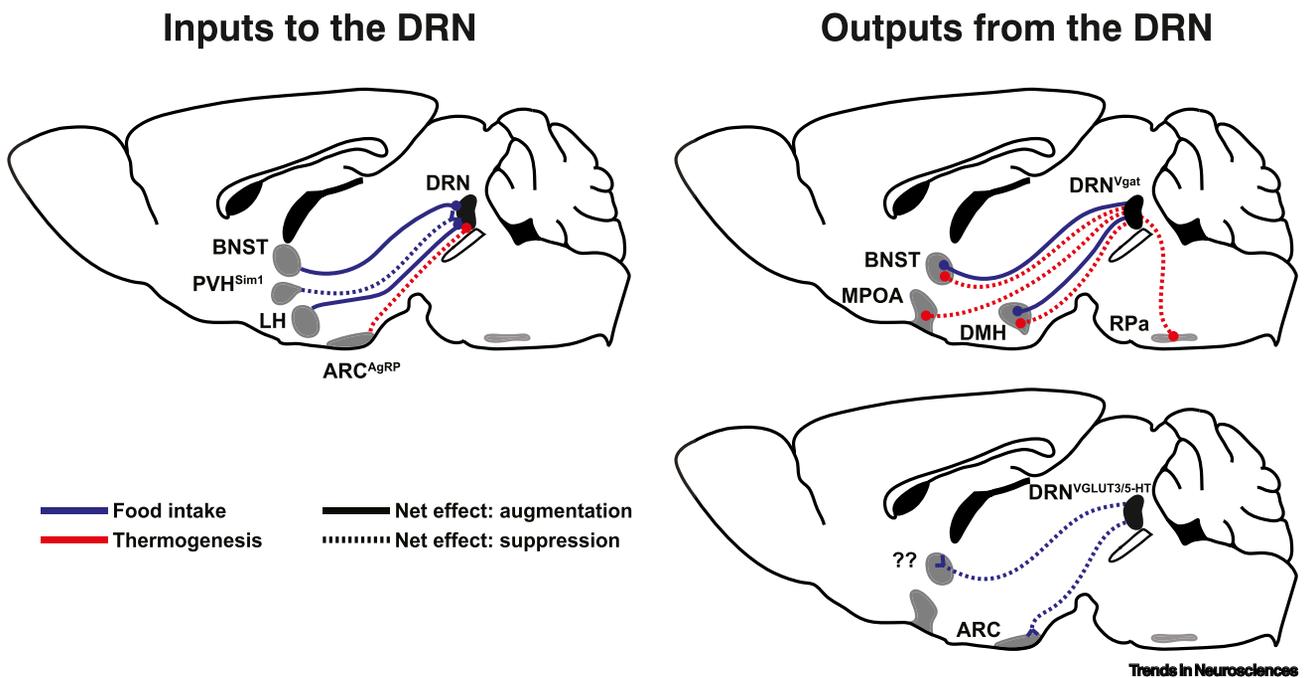
**Box 1. The role of the DRN in motivated behavior**

DRN neurons are known to play an important role in motivated behavior, and the existence of DRN circuits with overlapping roles in energy homeostasis and reward could relate to the drive to seek out and consume food. Notably, the VTA – including VTA DA neurons classically implicated in reward processing – receives input from serotonergic, glutamatergic, and GABAergic DRN neurons that is substantially greater than input from other loci (e.g., CeA, BNST, PVH, DMH, lateral PB) implicated in energy homeostasis. Indeed, DRN neurons constitute one of the single largest sources of input to the VTA [17,27,121–124]. However, while electrophysiological [125] and pharmacological studies targeting DRN GABA/5-HT signaling or opioid receptors have long suggested a role for the DRN in reinforcement [126–128], the role of specific DRN cell types in motivated behavior remains somewhat unclear.

Several studies indicate that DRN<sup>5-HT</sup> neurons are activated by various rewards, including nutritive rewards, and during periods leading up to expected rewards [18,107,129–131]. However, there are conflicting reports on whether directly stimulating DRN<sup>5-HT</sup> neurons signals reward [18,105,132,133]. One hypothesis is that DRN<sup>5-HT</sup> activation, while not intrinsically rewarding, might promote waiting for delayed rewards [133,134]. Complicating this picture is the fact that there appears to be significant heterogeneity in the way DRN<sup>5-HT</sup> neurons are modulated by both rewarding and noxious stimuli, the latter having also been reported to elicit DRN<sup>5-HT</sup> activity [107,135,136].

While the precise role of the DRN<sup>5-HT</sup> population in reward processing remains unclear, recent work has demonstrated that both DRN<sup>VGLUT3</sup> and DRN<sup>5-HT</sup> neurons monosynaptically innervate nucleus accumbens (NAc)-projecting VTA<sup>DA</sup> neurons [124,137,138]. Consistent with these anatomic findings, stimulating specifically VTA-projecting DRN neurons – including DRN<sup>5-HT</sup> neurons, and the partly overlapping DRN<sup>VGLUT3</sup> neurons that collectively constitute a larger VTA-projecting population – is both reinforcing and promotes DA release in the NAc [18,124,132,137,138].

Considerably less attention has been paid to other DRN cell types in the context of reward processing. While activating DRN<sup>GABA</sup> neurons might be mildly reinforcing, these cells also appear to be inhibited by rewards or expected rewards or even activated by noxious stimuli [130,132]. Meanwhile, though stimulating DRN<sup>DA</sup> neurons is likely not reinforcing and might even be aversive [7,132], these neurons can respond to both rewarding and noxious stimuli (or conditioned stimuli). They also appear necessary for the expression, rather than formation, of reward- and punishment-related memories [139].



**Figure 1. Reported effects of dorsal raphe nucleus (DRN) efferent and afferent projections on energy balance.** DRN glutamatergic (DRN<sup>VGLUT3</sup>), GABAergic (DRN<sup>VGAT</sup>), and serotonergic (DRN<sup>5-HT</sup>) neurons are embedded within an extended circuit known to regulate energy homeostasis. Differentiable effects on food intake (blue) and thermogenesis (red) can be driven by modulating specific DRN inputs or outputs. Abbreviations: AgRP, agouti-related peptide; ARC, arcuate nucleus of the hypothalamus; BNST, bed nucleus of the stria terminalis; DMH, dorsomedial hypothalamus; LH, lateral hypothalamus; MPOA, medial preoptic area; PVH, paraventricular nucleus of the hypothalamus; RPa, raphe pallidus; SIM1, single-minded homolog 1.

AgRP neurons leads to increased expression of cFos – a surrogate marker gene for neural activity – in the DRN [29,30]. While several studies have found that activation of AgRP neurons directly projecting to the DRN is insufficient to modulate food intake [29–31], it remains possible that polysynaptic interactions between AgRP and DRN neurons may augment feeding.

DRN neurons are also innervated by single-minded homolog 1 (SIM1)-expressing neurons in the paraventricular nucleus of the hypothalamus (PVH), which themselves receive extensive input from AgRP neurons [29,32]. While activating AgRP terminals in the DRN has little effect on feeding, silencing PVH<sup>SIM1</sup> axons in the DRN/vIPAG acutely increases food intake (Figure 1) [33]. Notably, optogenetic stimulation of PVH neurons expressing the melanocortin 4 receptor (MC4R) that project to the DRN/vIPAG, which represent a subset of PVH<sup>SIM1</sup> neurons, does not alter feeding behavior [34]. These studies together present a complex circuit logic through which the hypothalamus and DRN interact to drive changes in feeding.

The DRN also expresses receptors for peripheral molecules known to regulate feeding. Included in this group are receptors for ghrelin (GHSR) [35] and leptin (LEPR) [36]. Ghrelin injection into the DRN increases food intake and modulates the activity of DRN neurons [23,37,38]. However, there is little evidence that leptin-mediated signaling in DRN<sup>5-HT</sup> neurons affects energy intake. Despite an earlier report suggesting that serotonin-releasing DRN neurons respond to leptin via LEPRb [39], leptin-responsive and LEPRb-expressing DRN neurons appear to be largely nonserotonergic. Concordantly, depleting central serotonin does not alter the anorectic effects of leptin administration, nor does inactivating LEPRb in 5-HT neurons alter body weight or adiposity [40].

Pharmacological studies have suggested that, in addition to leptin and ghrelin, several peptide systems can operate in the DRN to affect energy homeostasis. For example, agonizing  $\alpha$ -2 adrenergic receptors in the DRN can increase food intake [41]. In contrast, activating glucagon-like peptide-1 (GLP-1) receptors expressed in the DRN reduces food intake and body weight [42,43], and increased neuromedin U (NMU) or estradiol signaling in the DRN likewise appears to decrease food intake [44,45]. Molecular profiling and histological studies have found that subsets of DRN neurons express additional neuropeptides or neuropeptide receptors implicated in regulating energy homeostasis (Box 2).

While studies have demonstrated impressive molecular heterogeneity among DRN neurons, the functional significance of these neuropeptide systems and their signaling in the DRN remains largely unknown. Despite a limited understanding of the function of specific neuropeptides, converging lines of evidence have established a clear role for the more broadly defined cell types in DRN – including those releasing serotonin, glutamate, and GABA – in the context of feeding (Table 1).

Central 5-HT has been known to play a role in feeding for many decades [46]. However, establishing a definitive link between specific serotonin-releasing neurons and feeding has often remained elusive. Toward this end, numerous groups have sought to molecularly and neuroanatomically characterize 5-HT neurons. Recent studies using rabies virus-based tracing, for example, have found that DRN<sup>5-HT</sup> neurons receive significant monosynaptic input from loci implicated in energy balance, such as the CeA, BNST, LH, PVH, tuberomammillary nucleus (TMN), dorsomedial hypothalamus (DMH), and PBN – including from projection neurons that express peptides such as orexin, melanin-concentrating hormone (MCH), and vasopressin [17,25–28]. DRN<sup>5-HT</sup> neurons, in turn, project directly back to many of these regions [17].

### Box 2. Neuropeptide signaling in the DRN

Numerous neuropeptides are critical in the central control of energy balance. While the DRN contains several classes of peptidergic neurons and neuropeptide receptors (see also main text), the functional role of many peptidergic systems, particularly within the DRN, is not well understood. Nonetheless, the remarkable mosaic of DRN cell types – beyond just 5-HT, DA, VGLUT3, and GABA neurons – suggests intriguing hypotheses about the DRN subpopulations that may be involved in energy homeostasis. It is important to note, however, that while the expression pattern of DRN<sup>5-HT</sup> neurons is broadly conserved across mammals, neuropeptide expression in the DRN can display significant cross-species variation [16].

Terminals from several types of peptidergic neurons are distributed throughout the DRN [16], and DRN neurons are known to express many post-synaptic receptors responsive to these neuropeptide classes. Examples include corticotropin-releasing factor receptors [140,141], orexin receptors [142,143], neurotensin receptors [144,145], mu/kappa opioid receptors [11,128], galanin receptors [146,147], and the neurokinin 1 receptor [148]. Histologic and electrophysiologic studies have demonstrated that these receptors all colocalize, at least to some extent, with DRN<sup>5-HT</sup> neurons.

In terms of efferent signaling, DRN neurons express several peptides linked to energy balance. These include substance P [149,150], cholecystokinin, vasoactive intestinal peptide (VIP) [16,151], several opioid peptide hormones [16,152], neurotensin [16], galanin [147], and cocaine/amphetamine-regulated transcript [153]. In particular, substance P is known to be expressed in DRN<sup>5-HT</sup> neurons, while VIP is expressed in DRN<sup>DA</sup> neurons.

Relatively few studies have examined neuropeptide and neuropeptide receptor expression in specific DRN cell types beyond DRN<sup>5-HT</sup> neurons. However, some peptides associated with energy homeostasis are found in the lateral aspects of DRN where GABAergic neurons are abundant. These include neuropeptide Y [16,154] and thyrotropin-releasing hormone [150,155].

More recently, single-cell RNA sequencing has provided additional insight into the DRN's molecular diversity while solidifying previous findings [19,48,49]. These studies have helped confirm, for instance, that DRN<sup>GABA</sup> neurons express somatostatin (SST) [16,150] while also establishing that they express prepronociceptin (PNOC) [19]. Similarly, single-cell profiling of DRN<sup>5-HT</sup> neurons has shown that subsets of these neurons express, in addition to the peptides and receptors listed above, corticotropin-releasing hormone, growth hormone-releasing hormone, and receptors for both histamine and calcitonin [19,48,49].

Much of the functional evidence linking DRN<sup>5-HT</sup> neurons to feeding comes from studies that pharmacologically modulate the autoinhibitory 5-HT<sub>1A</sub> receptor, which is robustly expressed across DRN<sup>5-HT</sup> cells [47–49]. 5-HT<sub>1A</sub> receptor agonists are known to suppress activity in DRN<sup>5-HT</sup> neurons, which is similar to the autoinhibitory effect observed when 5-HT is applied to these cells [50]. The most frequently studied 5-HT<sub>1A</sub> agonist has been 8-OH-DPAT, which is selective [51] and has been shown to both hyperpolarize DRN<sup>5-HT</sup> neurons and ultimately reduce 5-HT release [50,52].

Pharmacological studies have demonstrated that 8-OH-DPAT microinjection in the DRN of sated rodents increases feeding [23,53–57]. Notably, the orexigenic effect observed when inhibiting DRN 5-HT release is consistent with the anorectic effects of 5-HT signaling in the ARC [58–60]. It also accords with the orexigenic effects of systemic 8-OH-DPAT administration [61,62] and administration of 5-HT or agents that enhance 5-HT bioavailability (e.g., fenfluramine) within the DRN [53].

One limitation of earlier pharmacological studies has been the predominant use of male rodents; recent work has suggested that adult-onset disruption of DRN 5-HT synthesis promotes hyperphagia and weight gain in a sex-dependent manner [63]. Furthermore, 8-OH-DPAT might have distinct effects on feeding during the dark cycle or in starved animals [64,65]. Still, the balance of evidence indicates that reduced 5-HT release from the DRN, secondary to 5-HT<sub>1A</sub> receptor activation, increases food intake.

Although the mechanism through which specific DRN<sup>5-HT</sup> projections regulate energy intake remains poorly understood, one recent study found that photoactivation of DRN<sup>5-HT</sup> neurons

projecting to the ARC reduces feeding (Figure 1) [66]. This result is consistent with the fact that ARC-projecting DRN<sup>5-HT</sup> neurons innervate both classically anorectic POMC neurons (which express excitatory 5-HT<sub>2C</sub> receptors) and orexigenic AGRP neurons (which express inhibitory 5-HT<sub>1B</sub> receptors) [66,67].

Previous studies have shown that serotonin exhibits anorectic effects by acting at 5-HT<sub>2C</sub> receptors expressed in POMC neurons. In addition to ARC<sup>POMC</sup> neurons, 5-HT<sub>2C</sub> receptors are found in POMC neurons located in the NTS. Signaling at 5-HT<sub>2C</sub> receptors in both the ARC and NTS reduces food intake and helps maintain normal body weight and glucose homeostasis [58–60,68,69]. Indeed, the antiobesity medication and selective 5-HT<sub>2C</sub> receptor agonist lorcaserin [70] suppresses appetite by acting in the NTS and ARC on POMC neurons [69], which also mediate the anorectic effects of the serotonin-releasing agent fenfluramine [59,60]. Additional work has suggested a functional role for serotonin signaling on AGRP neurons, through 5-HT<sub>1B</sub> receptors, as well [71].

Curiously, despite decades of evidence supporting an anorectic role for 5-HT release from the DRN, only one publication [66], to our knowledge, has described a DRN<sup>5-HT</sup> subpopulation directly suppressing food intake. Another recent report suggested, though, that chemogenetic modulation of DRN<sup>5-HT</sup> neurons does not affect feeding [31]. Functional heterogeneity among 5-HT neurons, as well as suboptimal recombinase driver lines targeting these populations, could help explain this discrepancy in the literature. For example, while DRN<sup>5-HT</sup> neurons are activated during refeeding, they can be activated during eating bouts in the sated state as well, and subsets of DRN<sup>5-HT</sup> neurons projecting to the ventral tegmental area (VTA) appear to regulate hedonic, rather than homeostatic, feeding [66]. Studies using enhanced intersectional genetic approaches could shed additional light on this heterogeneity. Of note, heterogeneity of 5-HT neurons appears to be conserved across many species, including *Drosophila*, where increased feeding can be driven by a small subset of 5-HT neurons within a broader population that, *in toto*, drives the opposite effect [72].

While the precise role of DRN<sup>5-HT</sup> neurons in feeding is only partly understood, VGLUT3-expressing neurons in the DRN also appear to have a clear role in regulating energy intake. This is notable, given that DRN<sup>VGLUT3</sup> neurons have considerable, though not complete, overlap with DRN<sup>5-HT</sup> neurons [15,17,73]. Furthermore, some DRN<sup>5-HT</sup> neurons are known to corelease glutamate [74], and VGLUT3 in axon terminals could potentially facilitate increased 5-HT release through vesicular synergy [75].

DRN<sup>VGLUT3</sup> neurons appear to be activated in refed mice, compared with fed mice, as measured by cFos expression [23,76]. Consistent with these findings, optogenetic or chemogenetic activation of DRN<sup>VGLUT3</sup> neurons decreases food intake in both normal and leptin-deficient obese mice, whereas photoinhibition of DRN<sup>VGLUT3</sup> neurons increases feeding. The latter effect is consistent with the notion that inhibiting DRN<sup>5-HT</sup> neurons (or specific subsets of them) can increase feeding. Indeed, DRN<sup>VGLUT3</sup> neurons express the 5-HT<sub>1A</sub> receptor and can be hyperpolarized by 8-OH-DPAT [23].

Despite the findings described above, it remains unknown whether there is a DRN subpopulation that is both necessary and sufficient to drive the effects of modulating serotonergic or glutamatergic DRN neurons, as a whole, on energy intake. One initial finding is that agonizing the neuropeptide Y receptor Y2 (NPY2R), which is expressed in both DRN<sup>VGLUT3</sup> and DRN<sup>5-HT</sup> neurons, can activate DRN<sup>VGLUT3</sup> neurons and decrease food intake [23,48,49]. This, however, appears to be only the beginning of the story: Recent single-cell RNA sequencing has uncovered clusters of DRN<sup>5-HT</sup>

neurons expressing numerous other neuropeptide and neuropeptide receptor genes implicated in energy balance [19,48,49] (Box 2).

While decades of pharmacological studies have demonstrated that DRN<sup>5-HT</sup> signaling plays a role in feeding, the importance of GABAergic populations in the DRN has emerged only recently [23]. Although these DRN<sup>GABA</sup> neurons are somewhat anatomically intermingled with DRN<sup>5-HT</sup> neurons, they are predominantly found in the lateral aspects of the DRN, and GABA/5-HT marker genes are generally not coexpressed [15,17,73]. DRN<sup>GABA</sup> and DRN<sup>5-HT</sup> neurons are thus more functionally distinguishable populations than DRN<sup>5-HT</sup> and DRN<sup>VGLUT3</sup> neurons, though there is some minimal overlap between the two populations [16,19,48,49].

Similar to DRN<sup>5-HT</sup> neurons, inhibitory neurons in the DRN and vIPAG receive monosynaptic input from loci implicated in energy balance, including the CeA, BNST, LH, PVH, lateral PBN, and ARC [24,25,28]. However, the functional role of DRN<sup>GABA</sup> neurons contrasts starkly with DRN<sup>5-HT</sup> and DRN<sup>VGLUT3</sup> neurons. DRN<sup>GABA</sup> neurons show significant cFos induction following overnight fasting [23]. Consistent with this result, optogenetic or chemogenetic stimulation of DRN<sup>GABA</sup> neurons increases food intake in both fed and fasted mice, while inhibiting DRN<sup>GABA</sup> neurons decreases food intake in fasted mice. The increase in feeding from stimulating DRN<sup>GABA</sup> somata can be replicated by photoactivating DRN<sup>GABA</sup> neurons that make ascending projections directly to the BNST or DMH (Figure 1) [77]. Moreover, chronically inhibiting DRN<sup>GABA</sup> neurons can reduce body weight in leptin-deficient obese mice [23].

Although heterogeneity among DRN<sup>GABA</sup> neurons has only begun to be explored, it appears that MC4R is expressed in a subset of DRN<sup>GABA</sup> neurons. Both chemogenetically inhibiting DRN<sup>MC4R</sup> neurons and inhibiting them by infusing the MC4R agonist  $\alpha$ -MSH into the DRN decreases food intake [23,78]. Furthermore, attenuating the inactivation and degradation of  $\alpha$ -MSH in the DRN – and thereby magnifying  $\alpha$ -MSH signaling – can likewise reduce food intake and body weight [78]. Despite the consistency of these results with the feeding effects driven by DRN<sup>GABA</sup> neurons,  $\alpha$ -MSH has also been reported to depolarize DRN<sup>MC4R</sup> neurons [31]. Moreover, the same report found that manipulation of DRN<sup>MC4R</sup> signaling had no effect on food intake [31], highlighting the need for deeper examination of the functional roles of this cell population.

Another caveat to the ascribed roles of DRN GABA neurons as discussed above is the observation that another, likely overlapping, population of GABAergic vIPAG neurons shows decreased activity upon feeding [24]. Photostimulating these neurons also reduces feeding, while acutely inhibiting them increases feeding. Furthermore, chronic stimulation or inhibition of vIPAG<sup>GABA</sup> neurons can alter body weight. These contrasting findings raise the possibility that cell subtypes within the coarsely classified DRN/vIPAG GABAergic population can drive opposing effects on energy intake, perhaps through recurrent (i.e., local GABA–GABA) interactions. Regardless, these findings demonstrate a previously underappreciated role for DRN<sup>GABA</sup> neurons in regulating feeding.

### The role of the DRN in energy expenditure

Converging lines of evidence support an important role specifically for the DRN, particularly GABAergic and 5-HT neurons, in thermoregulation. Moreover, evidence suggests a role for these neurons in the control of energy expenditure through not only brown adipose tissue (BAT) regulation but also behavioral mechanisms extending beyond thermoregulation.

Early electrophysiological studies reported on DRN neurons whose firing rates are modulated by changes in local or ambient temperature [79,80]. Additional studies found that electrical

stimulation of the DRN can alter interscapular brown adipose tissue (iBAT) thermogenesis and concomitantly core temperature [81]. Pharmacological manipulations, such as infusing morphine or muscimol in the DRN, have also been shown to alter core body temperature [82,83].

Studies of the DRN's afferent and efferent connectivity have further supported a role for the DRN, particularly DRN<sup>GABA</sup> neurons, in energy expenditure. For instance, the DRN and vPAG receive significant input from the medial preoptic area (MPOA), which is implicated in thermoregulation; among these inputs are the BDNF/PACAP-expressing, warm-sensitive neurons that directly regulate body temperature [25,28,84]. Furthermore, photostimulation or chemogenetic activation of DRN-projecting ARC<sup>AgRP</sup> neurons decreases iBAT thermogenesis and core temperature, while chemical ablation of these neurons increases energy expenditure [31]. DRN<sup>GABA</sup> neurons, in particular, also make ascending, reciprocal projections to brain regions implicated in thermogenesis, such as the MPOA, LH, and PBN [77].

In addition, DRN and vPAG neurons – including DRN<sup>GABA</sup> neurons, and more specifically DRN<sup>GABA</sup> neurons that are activated by ambient warmth – polysynaptically innervate iBAT via a monosynaptic projection to the raphe pallidus (RPa), which is a major site of sympathetic outflow to the periphery [77,85–87]. Notably, DRN neurons receive afferent projections from iBAT [85,86], suggesting a rapid sensorimotor feedback loop between the DRN and iBAT, the function of which is largely unknown. These observations are consistent with evidence that sympathetic nerve activity to iBAT is increased by disinhibition of RPa neurons [88]. Interestingly, there also exist reciprocal connections between DRN neurons and inguinal white adipose tissue, suggesting a circuit mechanism by which the DRN might respond to changes in energy stores [86].

In addition to these anatomical observations, studies have demonstrated a functional role for DRN<sup>GABA</sup> neurons in controlling energy expenditure. Ambient warmth leads to increased activity in DRN<sup>GABA</sup> neurons, as measured through cFos induction [77]. Thermal challenge also augments cFos expression in the DRN as a whole, with variability in expression among serotonergic and nonserotonergic populations across subregions of the DRN [77,89,90]. Furthermore, chemogenetically stimulating DRN<sup>GABA</sup> neurons decreases iBAT thermogenesis, reduces the expression of thermogenesis-related genes in iBAT, and concomitantly lowers core body temperature and overall metabolic activity [77]. This suppression of iBAT thermogenesis can be recapitulated by projection-specific photoactivation of DRN<sup>GABA</sup> neurons innervating loci known to regulate thermogenesis (Figure 1). These loci include the DMH, BNST, and MPOA (ascending pathways) along with the RPa (descending pathway) [77]. In concordance with these findings, acute chemogenetic inhibition of DRN<sup>GABA</sup> neurons increases core body temperature and overall energy expenditure, likely through changes in locomotor activity [77].

Recent work has also illustrated an important but unclear role in energy expenditure for DRN<sup>MC4R</sup> neurons, subsets of which overlap with DRN<sup>GABA</sup> neurons and are projection targets for ARC<sup>AgRP</sup> neurons [31,78]. Chemogenetic inhibition or ablation of DRN<sup>MC4R</sup> neurons has been reported to decrease iBAT thermogenesis and core temperature, which accords with the inhibitory effect of AgRP on these cells [31]. In contrast, however, attenuating  $\alpha$ -MSH inactivation and degradation in the DRN – which likely chronically inhibits subsets of DRN<sup>MC4R</sup> and DRN<sup>GABA</sup> neurons – can augment iBAT thermogenesis marker gene expression, core body temperature, and metabolic activity [78]. Despite the discrepant effects on food intake and energy expenditure observed when modulating DRN<sup>MC4R</sup> neurons, this subpopulation could contribute to the increased energy expenditure thought to be driven by MC4R signaling outside of the PVH [91].

While the role of DRN<sup>5-HT</sup> neurons in energy expenditure has received relatively limited attention, recent work has reported that chemogenetic activation of DRN<sup>5-HT</sup> neurons can augment iBAT thermogenesis and chemogenetic inhibition suppresses it [31]. These findings are generally in line with observations from early pharmacological studies, which found that intra-DRN administration of agents that predominantly inhibit 5-HT neurons (5-HT, 8-OH-DPAT, or the nonselective 5-HT receptor agonist 5-CT) decreases core body temperature [82,92]. Systemic 8-OH-DPAT treatment elicits a similar hypothermic effect [93,94]. These results are also consistent with the more general and well-established finding that decreasing central 5-HT signaling – through lesions during development or adulthood – reduces core body temperature and BAT thermogenesis and leads to impaired adaptive responses to cold challenge [95–99]. Taken together, these results support the conclusion that 5-HT signaling in the DRN is necessary for maintaining normal energy expenditure.

Changes in mammalian energy expenditure can stem from changes in not only BAT thermogenesis but also locomotor activity [100]. Besides increasing energy expenditure directly, motor activity holds additional importance in the context of energy balance. For instance, increased locomotion can indicate foraging behavior and can thus represent a behavioral opportunity cost with respect to food intake. Indeed, a growing body of evidence suggests that neural populations in the DRN play a significant, though incompletely understood, role in locomotion.

Early studies reported that both muscimol infusion into the DRN [22] and nonspecifically photoactivating DRN neurons [9] can increase locomotion. There are also widespread, reciprocal projections between both serotonergic and nonserotonergic DRN neurons and nuclei within the basal ganglia, suggesting potential circuit mechanisms through which the DRN regulates locomotion [27,28,101]. Intriguingly, DRN<sup>5-HT</sup> neurons are also known to send collaterals into the spinal cord [102].

Additional studies have focused on the locomotor effects of modulating specific DRN cell types. While several pharmacological reports suggest that reducing DRN<sup>5-HT</sup> signaling can decrease locomotion [103,104], direct photoactivation of DRN<sup>5-HT</sup> neurons appears to suppress locomotion while not causing generalized motor impairment or an anxiety-like response [105]. Meanwhile, photoinhibition of DRN<sup>GABA</sup> neurons profoundly increases locomotor activity, and DRN<sup>GABA</sup> activation can either decrease locomotion [23,77] or increase it in specific behavioral contexts [106]. Importantly, thermogenesis studies under general anesthesia suggest that the core temperature changes caused by DRN<sup>GABA</sup> excitation and inhibition may depend, in part, on changes in locomotion [77].

Several plausible explanations exist for the discordant effects on locomotion driven by DRN neurons. First, it seems likely that specific DRN<sup>5-HT</sup> subpopulations can mediate different effects on locomotion. For example, while chemogenetic activation of DRN<sup>5-HT</sup> neurons projecting to the orbitofrontal cortex (OFC) or CeA decreases locomotion, photoactivation of BNST-projecting DRN<sup>5-HT</sup> neurons does not affect locomotion, and photoactivation of the overlapping DRN<sup>VGLUT3</sup> neurons in both normal-weight and obese mice increases locomotion [23,107,108].

A second possibility is that an animal's internal state modulates the relationship between DRN 5-HT or GABA signaling and locomotion. Movement can increase or decrease DRN<sup>5-HT</sup> neural activity, depending on whether an animal is in a 'low-threat' or 'high-threat' environment, and DRN<sup>5-HT</sup> photostimulation can lead to facilitation or suppression of locomotion in environments with different degrees of threat. Activity in DRN<sup>GABA</sup> neurons seems similarly modulated by environmental threat level [106].

Yet another hypothesis is that DRN neurons, particularly 5-HT neurons, play a greater role in the coordination of motor behaviors than locomotor activity *per se*. This is supported by electrophysiological studies indicating that subsets of DRN<sup>5-HT</sup> neurons are rapidly activated by specific movements, including grooming, licking, head motion, and chewing, in addition to responding to treadmill-induced locomotion [109,110]. Interestingly, DRN<sup>5-HT</sup> neural activity in nonmammals also can play a ‘coordinating’ role: for example, in calibrating locomotor drive to displacement of the body in the environment in zebrafish [111].

In addition to locomotion and BAT thermogenesis, energy expenditure is generally correlated with heart rate [112]. Some electrophysiological and pharmacological evidence suggests that altering DRN neural activity can affect both heart rate and blood pressure [113,114]. Other experiments using excitatory amino acid injections have suggested a role for the vPAG in modulating heart rate and blood pressure [115]. Despite these observations, it remains unclear what effect these fluctuations might have on overall metabolic rate and whether DRN neurons respond to other sympathetic parameters under physiologic conditions.

### Concluding remarks

The aggregate molecular, cellular, circuit, and functional data reviewed here support a key role for the DRN in the regulation of energy balance. These data also suggest that the actions of DRN<sup>GABA</sup> neurons generally oppose those of DRN<sup>5-HT</sup> and DRN<sup>VGLUT3</sup> neurons. Activity among DRN<sup>GABA</sup> neurons appears to signal a state of energy deficit in which an animal will consume food to restore depleted energy stores and expend less energy to retain those available. In contrast, activity in DRN<sup>5-HT</sup> and DRN<sup>VGLUT3</sup> neurons appears to signal a state of energy surfeit in which food intake is reduced and aggregate energy expenditure is increased (Table 1).

While there are multiple possible circuit-level explanations for this opponency, among the most straightforward ones is a model in which GABAergic inhibition of 5-HT or VGLUT3 neurons via local collaterals constitutes a dominant circuit within the DRN. Evidence supports the existence of such connectivity: Viral tracing has shown that DRN<sup>5-HT</sup> neurons are monosynaptically innervated by local GABAergic neurons [25], in addition to their innervation by other 5-HT neurons. Studies have also shown that photoinhibition or photostimulation of DRN<sup>GABA</sup> neurons can alter activity in neighboring DRN<sup>5-HT</sup> and DRN<sup>VGLUT3</sup> neurons [8,23,28].

Elaborating on the role of the DRN in energy homeostasis – and, ultimately, identifying appropriate druggable targets – will be facilitated by intersectional genetic approaches that can interrogate neural circuits with great precision. While it is clear that DRN neurons exert potent and bidirectional control over energy intake and expenditure, one key question is to what extent the DRN serotonergic, GABAergic, or glutamatergic populations that control energy intake overlap with (or are separate from) populations that regulate energy expenditure and motivated behavior (see Outstanding questions).

It is also unknown what minimal DRN subpopulations are necessary and sufficient to drive the effects on energy balance observed when coarsely modulating DRN cell types. These subpopulations might be demarcated by afferent or efferent projection targets, the expression of neuropeptides, neuropeptide receptors, or some combination of these. Such DRN subpopulations could drive effects that have not been observed when manipulating broader DRN cell types – for instance, it remains unclear whether there exists a DRN population that can augment iBAT thermogenesis. Theoretically, such a population might be found among the significant proportion of iBAT-projecting DRN neurons that are not GABAergic [77].

### Outstanding questions

How do the DRN serotonergic, GABAergic, and glutamatergic populations regulate energy balance? Are there individual populations that simultaneously coordinate energy intake and expenditure? Or are there independent, interacting cell subtypes responsible for controlling different aspects of energy balance?

What DRN subpopulations are necessary and sufficient to produce the effects on energy intake and expenditure from modulating the broad, principal DRN cell types? Can these neural subpopulations be delineated by molecular markers, projection patterns, or a combination of properties?

How are signals related to energy imbalance transmitted to the DRN from the periphery and other central loci? Are these signals neural, humoral, or both?

What are the short-term dynamics of DRN serotonergic, GABAergic, and glutamatergic neurons, as related to energy homeostasis (e.g., thermal challenge, food presentation)?

To what extent are different DRN output circuits necessary or redundant in producing effects on food intake and/or thermogenesis?

Further exploration of the functional role of neuropeptides and peptide receptors seems promising, given the fact that the neuropeptide and receptor composition of the DRN mirrors hypothalamic loci implicated in energy balance. For example, numerous peptides and receptors (e.g., LEPR, NPY, cocaine/amphetamine-regulated transcript, 5-HT<sub>2C</sub>, GHSR) demarcating ARC populations functionally implicated in energy homeostasis [116–119] are expressed throughout the DRN (Box 2). Similarly, subsets of DRN neurons, such as neurons in the PVH, express thyrotropin-releasing hormone and MC4R, which have well-established roles in energy balance [34,120]. These parallels suggest that interrogating peptide signaling could further clarify the DRN's role in energy homeostasis.

At a circuit level, the redundancy or necessity of specific DRN projection sets that regulate energy balance remains largely unknown, though it seems that different projections can regulate distinct aspects of energy balance [77]. It will thus be crucial to parse how much DRN neurons collateralize across projection targets, which would enable a richer understanding of how different DRN outputs coordinate energy balance. In particular, DRN<sup>GABA</sup> neurons could modulate energy intake and expenditure through a combination of local and long-range projections – which is intriguing, given the historic emphasis on local GABAergic interneurons. More broadly, variability in the observed effects driven by circuits in close proximity (e.g., DRN-projecting vs. vIPAG-projecting AgRP neurons, or DRN<sup>GABA</sup> vs. vIPAG<sup>GABA</sup> neurons) suggests a rich local architecture within broader molecularly defined DRN cell types [24,31].

Additional work is also needed to understand the mechanisms by which internal and external signals regulate DRN neural activity. Signals conveying energy imbalance may be transmitted to the DRN from a multitude of central and peripheral cell types, but which afferent circuits are functionally relevant is incompletely understood. Moreover, little attention has been paid to how the DRN encodes energy-related signals. Limited data exist on the real-time dynamics of DRN populations in response to relevant stimuli (e.g., food presentation in the hunger state, thermal challenge, or other states of physiological need). These data would complement results observed from causal circuit manipulations, which can be supraphysiologic. It is also possible that long-term shifts in peripheral physiology might alter the molecular profile of DRN subpopulations.

Extensive work on the hypothalamus has revealed cell types distributed across several nuclei that are involved in energy homeostasis. However, increased attention to loci outside the hypothalamus has established the importance of brainstem circuits in sensing and responding to changes in energy state. Although the DRN has a critical role within the extended circuit regulating energy homeostasis, further research may elucidate how energy-related signals are integrated in the DRN and the specific DRN subpopulations responsible for maintaining energy balance.

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### Declaration of interests

The authors declare no competing interests in relation to this work.

### References

1. Aponte, Y. *et al.* (2011) AgRP neurons are sufficient to orchestrate feeding behavior rapidly and without training. *Nat. Neurosci.* 14, 351–355
2. Shi, Y.C. *et al.* (2013) Arcuate NPY controls sympathetic output and BAT function via a relay of tyrosine hydroxylase neurons in the PVN. *Cell Metab.* 17, 236–248

3. Carter, M.E. *et al.* (2013) Genetic identification of a neural circuit that suppresses appetite. *Nature* 503, 111–114
4. Kim, D.Y. *et al.* (2020) A neural circuit mechanism for mechanosensory feedback control of ingestion. *Nature* 580, 376–380
5. Wu, Q. *et al.* (2012) Deciphering a neuronal circuit that mediates appetite. *Nature* 483, 594–597
6. D'Agostino, G. *et al.* (2016) Appetite controlled by a cholecystokinin nucleus of the solitary tract to hypothalamus neurocircuit. *Elife* 5, e12225
7. Matthews, G.A. *et al.* (2016) Dorsal raphe dopamine neurons represent the experience of social isolation. *Cell* 164, 617–631
8. Challis, C. *et al.* (2013) Raphe GABAergic neurons mediate the acquisition of avoidance after social defeat. *J. Neurosci.* 33, 13978–13988
9. Warden, M.R. *et al.* (2012) A prefrontal cortex-brainstem neuronal projection that controls response to behavioural challenge. *Nature* 492, 428–432
10. Teissier, A. *et al.* (2015) Activity of raphe serotonergic neurons controls emotional behaviors. *Cell Rep.* 13, 1965–1976
11. Li, C. *et al.* (2016) Mu opioid receptor modulation of dopamine neurons in the periaqueductal gray/dorsal raphe: a role in regulation of pain. *Neuropsychopharmacology* 41, 2122–2132
12. Cho, J.R. *et al.* (2017) Dorsal raphe dopamine neurons modulate arousal and promote wakefulness by salient stimuli. *Neuron* 94, 1205–1219
13. Okaty, B.W. *et al.* (2015) Multi-scale molecular deconstruction of the serotonin neuron system. *Neuron* 88, 774–791
14. Ishimura, K. *et al.* (1988) Quantitative analysis of the distribution of serotonin-immunoreactive cell bodies in the mouse brain. *Neurosci. Lett.* 91, 265–270
15. Hioki, H. *et al.* (2010) Vesicular glutamate transporter 3-expressing nonserotonergic projection neurons constitute a subregion in the rat midbrain raphe nuclei. *J. Comp. Neurol.* 518, 668–686
16. Fu, W. *et al.* (2010) Chemical neuroanatomy of the dorsal raphe nucleus and adjacent structures of the mouse brain. *J. Comp. Neurol.* 518, 3464–3494
17. Cardozo Pinto, D.F. *et al.* (2019) Characterization of transgenic mouse models targeting neuromodulatory systems reveals organizational principles of the dorsal raphe. *Nat. Commun.* 10, 4633
18. Liu, Z. *et al.* (2014) Dorsal raphe neurons signal reward through 5-HT and glutamate. *Neuron* 81, 1360–1374
19. Huang, K.W. *et al.* (2019) Molecular and anatomical organization of the dorsal raphe nucleus. *Elife* 8, e46464
20. Wyler, S.C. *et al.* (2017) Serotonergic control of metabolic homeostasis. *Front. Cell. Neurosci.* 11, 277
21. Coscina, D.V. and Stancer, H.C. (1977) Selective blockade of hypothalamic hyperphagia and obesity in rats by serotonin-depleting midbrain lesions. *Science* 195, 416–419
22. Przewlocka, B. *et al.* (1979) Evidence that GABA in the nucleus dorsalis raphe induces stimulation of locomotor activity and eating behavior. *Life Sci.* 25, 937–945
23. Nectow, A.R. *et al.* (2017) Identification of a brainstem circuit controlling feeding. *Cell* 170, 429–442
24. Hao, S. *et al.* (2019) The lateral hypothalamic and BNST GABAergic projections to the anterior ventrolateral periaqueductal gray regulate feeding. *Cell Rep.* 28, 616–624
25. Weissbourd, B. *et al.* (2014) Presynaptic partners of dorsal raphe serotonergic and GABAergic neurons. *Neuron* 83, 645–662
26. Dorocic, I.P. *et al.* (2014) A whole-brain atlas of inputs to serotonergic neurons of the dorsal and median raphe nuclei. *Neuron* 83, 663–678
27. Ogawa, S.K. *et al.* (2014) Organization of monosynaptic inputs to the serotonin and dopamine neuromodulatory systems. *Cell Rep.* 8, 1105–1118
28. Zhou, L. *et al.* (2017) Organization of functional long-range circuits controlling the activity of serotonergic neurons in the dorsal raphe nucleus. *Cell Rep.* 18, 3018–3032
29. Betley, J.N. *et al.* (2013) Parallel, redundant circuit organization for homeostatic control of feeding behavior. *Cell* 155, 1337–1350
30. Steculorum, S.M. *et al.* (2016) AgRP neurons control systemic insulin sensitivity via myostatin expression in brown adipose tissue. *Cell* 165, 125–138
31. Han, Y. *et al.* (2021) Deciphering an AgRP-serotonergic neural circuit in distinct control of energy metabolism from feeding. *Nat. Commun.* 12, 3525
32. Geerling, J.C. *et al.* (2010) Paraventricular hypothalamic nucleus: axonal projections to the brainstem. *J. Comp. Neurol.* 518, 1460–1499
33. Stachniak, T.J. *et al.* (2014) Chemogenetic synaptic silencing of neural circuits localizes a hypothalamus→midbrain pathway for feeding behavior. *Neuron* 82, 797–808
34. Garfield, A.S. *et al.* (2015) A neural basis for melanocortin-4 receptor-regulated appetite. *Nat. Neurosci.* 18, 863–871
35. Zigman, J.M. *et al.* (2006) Expression of ghrelin receptor mRNA in the rat and the mouse brain. *J. Comp. Neurol.* 494, 528–548
36. Scott, M.M. *et al.* (2009) Leptin targets in the mouse brain. *J. Comp. Neurol.* 514, 518–532
37. Carlini, V.P. (2004) Differential role of the hippocampus, amygdala, and dorsal raphe nucleus in regulating feeding, memory, and anxiety-like behavioral responses to ghrelin. *Biochem. Biophys. Res. Commun.* 313, 635–641
38. Hansson, C. *et al.* (2011) Central administration of ghrelin alters emotional responses in rats: behavioural, electrophysiological and molecular evidence. *Neuroscience* 180, 201–211
39. Yadav, V.K. (2009) A serotonin-dependent mechanism explains the leptin regulation of bone mass, appetite, and energy expenditure. *Cell* 138, 976–989
40. Lam, D.D. (2011) Leptin does not directly affect CNS serotonin neurons to influence appetite. *Cell Metab.* 13, 584–591
41. Flores, R.A. (2021) Injections of the  $\alpha$ -2 adrenoceptor agonist clonidine into the dorsal raphe nucleus increases food intake in satiated rats. *Neuropharmacology* 182, 108397
42. Merchenthaler, I. *et al.* (1999) Distribution of pre-pro-glucagon and glucagon-like peptide-1 receptor messenger RNAs in the rat central nervous system. *J. Comp. Neurol.* 403, 261–280
43. Anderberg, R.H. *et al.* (2017) Glucagon-like peptide 1 and its analogs act in the dorsal raphe and modulate central serotonin to reduce appetite and body weight. *Diabetes* 66, 1062–1073
44. McCue, D.L. *et al.* (2017) Regulation of motivation for food by neuromedin U in the paraventricular nucleus and the dorsal raphe nucleus. *Int. J. Obes. (Lond)* 41, 120–128
45. Santollo, J. *et al.* (2011) Estradiol acts in the medial preoptic area, arcuate nucleus, and dorsal raphe nucleus to reduce food intake in ovariectomized rats. *Horm. Behav.* 60, 86–93
46. Saller, C.F. and Stricker, E.M. (1976) Hyperphagia and increased growth in rats after intraventricular injection of 5,7-dihydroxytryptamine. *Science* 192, 385–387
47. Day, H.E. *et al.* (2004) Differential expression of 5HT-1A, alpha 1b adrenergic, CRF-R1, and CRF-R2 receptor mRNA in serotonergic, gamma-aminobutyric acidergic, and catecholaminergic cells of the rat dorsal raphe nucleus. *J. Comp. Neurol.* 474, 364–378
48. Okaty, B.W. *et al.* (2020) A single-cell transcriptomic and anatomic atlas of mouse dorsal raphe *Pet1* neurons. *Elife* 9, e55523
49. Ren, J. *et al.* (2019) Single-cell transcriptomes and whole-brain projections of serotonin neurons in the mouse dorsal and median raphe nuclei. *Elife* 8, e49424
50. Sprouse, J.S. and Aghajanian, G.K. (1986) (-)-Propranolol blocks the inhibition of serotonergic dorsal raphe cell firing by 5-HT<sub>1A</sub> selective agonists. *Eur. J. Pharmacol.* 128, 295–298
51. Middlemiss, D.N. and Fozard, J.R. (1983) 8-Hydroxy-2-(di-n-propylamino)-tetralin discriminates between subtypes of the 5-HT<sub>1</sub> recognition site. *Eur. J. Pharmacol.* 90, 151–153
52. Hopwood, S.E. and Stamford, J.A. (2001) Multiple 5-HT (1) autoreceptor subtypes govern serotonin release in dorsal and median raphe nuclei. *Neuropharmacology* 40, 508–519
53. Fletcher, P.J. and Davies, M. (1990) Dorsal raphe microinjection of 5-HT and indirect 5-HT agonists induces feeding in rats. *Eur. J. Pharmacol.* 184, 265–271
54. Fletcher, P.J. (1991) Dopamine receptor blockade in nucleus accumbens or caudate nucleus differentially affects feeding induced by 8-OH-DPAT injected into dorsal or median raphe. *Brain Res.* 552, 181–189

55. Fletcher, P.J. and Coscina, D.V. (1993) Injecting 5-HT into the PVN does not prevent feeding induced by injecting 8-OH-DPAT into the raphe. *Pharmacol. Biochem. Behav.* 46, 487–491
56. Currie, P.J. *et al.* (1994) Administration of 8-OH-DPAT into the midbrain raphe nuclei: effects on medial hypothalamic NE-induced feeding. *Am. J. Phys.* 266, 1645–1651
57. Coscina, D.V. *et al.* (2000) Posterodorsal amygdala lesions reduce feeding stimulated by 8-OH-DPAT. *Brain Res.* 883, 243–249
58. Heisler, L.K. *et al.* (2002) Activation of central melanocortin pathways by fenfluramine. *Science* 297, 609–611
59. Xu, Y. *et al.* (2008) 5-HT<sub>2</sub>CRs expressed by pro-opiomelanocortin neurons regulate energy homeostasis. *Neuron* 60, 582–589
60. Xu, Y. *et al.* (2010) A serotonin and melanocortin circuit mediates D-fenfluramine anorexia. *J. Neurosci.* 30, 14630–14634
61. Dourish, C.T. *et al.* (1985) Low doses of the putative serotonin agonist 8-hydroxy-2-(di-*n*-propylamino) tetralin (8-OH-DPAT) elicit feeding in the rat. *Psychopharmacology* 86, 197–204
62. Bendotti, C. and Samanin, R. (1986) 8-Hydroxy-2-(di-*n*-propylamino) tetralin (8-OH-DPAT) elicits eating in free-feeding rats by acting on central serotonin neurons. *Eur. J. Pharmacol.* 121, 147–150
63. Liu, H. *et al.* (2021) TPH2 in the dorsal raphe nuclei regulates energy balance in a sex-dependent manner. *Endocrinology* 162, bqaa183
64. Currie, P.J. and Coscina, D.V. (1993) Diurnal variations in the feeding response to 8-OH-DPAT injected into the dorsal or median raphe. *Neuroreport* 4, 1105–1117
65. Ebenezer, I.S. (1992) Effects of the 5-HT<sub>1A</sub> agonist 8-OH-DPAT on food intake in food-deprived rats. *Neuroreport* 3, 1019–1022
66. He, Y. *et al.* (2021) 5-HT recruits distinct neurocircuits to inhibit hunger-driven and non-hunger-driven feeding. *Mol. Psychiatry* Published online July 21, 2021. <https://doi.org/10.1038/s41380-021-01220-z>
67. Wang, D. *et al.* (2015) Whole-brain mapping of the direct inputs and axonal projections of POMC and AgRP neurons. *Front. Neuroanat.* 9, 40
68. Berglund, E.D. *et al.* (2013) Serotonin 2C receptors in pro-opiomelanocortin neurons regulate energy and glucose homeostasis. *J. Clin. Invest.* 123, 5061–5070
69. D'Agostino, G. *et al.* (2018) Nucleus of the solitary tract serotonin 5-HT<sub>2C</sub> receptors modulate food intake. *Cell Metab.* 28, 619–630
70. Smith, S.R. *et al.* (2010) Multicenter, placebo-controlled trial of lorcaserin for weight management. *N. Engl. J. Med.* 363, 245–256
71. Heisler, L.K. *et al.* (2006) Serotonin reciprocally regulates melanocortin neurons to modulate food intake. *Neuron* 51, 239–249
72. Albin, S.D. *et al.* (2015) A subset of serotonergic neurons evokes hunger in adult *Drosophila*. *Curr. Biol.* 25, 2435–2440
73. Calizo, L.H. *et al.* (2011) Raphe serotonin neurons are not homogenous: electrophysiological, morphological and neurochemical evidence. *Neuropharmacology* 61, 524–543
74. Sengupta, A. *et al.* (2017) Control of amygdala circuits by 5-HT neurons via 5-HT and glutamate cotransmission. *J. Neurosci.* 37, 1785–1796
75. El Mestikawy, S. *et al.* (2011) From glutamate co-release to vesicular synergy: vesicular glutamate transporters. *Nat. Rev. Neurosci.* 12, 204–216
76. Wu, Q. *et al.* (2014) The temporal pattern of cfos activation in hypothalamic, cortical, and brainstem nuclei in response to fasting and refeeding in male mice. *Endocrinology* 155, 840–853
77. Schneeberger, M. *et al.* (2019) Regulation of energy expenditure by brainstem GABA neurons. *Cell* 178, 672–685
78. Bruschetta, G. *et al.* (2020) MC<sub>4</sub>R signaling in dorsal raphe nucleus controls feeding, anxiety, and depression. *Cell Rep.* 33, 108267
79. Weiss, B.L. and Aghajanian, G.K. (1971) Activation of brain serotonin metabolism by heat: role of midbrain raphe neurons. *Brain Res.* 26, 37–48
80. Cronin, M.J. and Baker, M.A. (1977) Thermosensitive midbrain neurons in the cat. *Brain Res.* 128, 461–472
81. Dib, B. *et al.* (1994) Thermogenesis in brown adipose tissue is activated by electrical stimulation of the rat dorsal raphe nucleus. *Brain Res.* 650, 149–152
82. Higgins, G.A. *et al.* (1988) Behavioural and biochemical consequences following activation of 5HT<sub>1</sub>-like and GABA receptors in the dorsal raphe nucleus of the rat. *Neuropharmacology* 27, 993–1001
83. Shen, Z. *et al.* (1986) Effect of periaqueductal morphine injection on thermal response in rats. *Jpn. J. Physiol.* 36, 485–496
84. Tan, C.L. *et al.* (2016) Warm-sensitive neurons that control body temperature. *Cell* 167, 47–59
85. Ryu, V. *et al.* (2015) Brown adipose tissue has sympathetic-sensory feedback circuits. *J. Neurosci.* 35, 2181–2190
86. Ryu, V. *et al.* (2017) Bidirectional crosstalk between the sensory and sympathetic motor systems innervating brown and white adipose tissue in male Siberian hamsters. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 312, R324–R337
87. Cano, G. *et al.* (2003) Anatomical substrates for the central control of sympathetic outflow to interscapular adipose tissue during cold exposure. *J. Comp. Neurol.* 460, 303–326
88. Morrison, S.F. *et al.* (1999) GABA-mediated inhibition of raphe pallidus neurons regulates sympathetic outflow to brown adipose tissue. *Am. J. Phys.* 276, R290–R297
89. Hale, M.W. *et al.* (2011) Evidence for in vivo thermosensitivity of serotonergic neurons in the rat dorsal raphe nucleus and raphe pallidus nucleus implicated in thermoregulatory cooling. *Exp. Neurol.* 227, 264–278
90. Bratincsák, A. and Palkovits, M. (2004) Activation of brain areas in rat following warm and cold ambient exposure. *Neuroscience* 127, 385–397
91. Balthasar, N. *et al.* (2005) Divergence of melanocortin pathways in the control of food intake and energy expenditure. *Cell* 123, 493–505
92. Hillegaart, V. (1991) Effects of local application of 5-HT and 8-OH-DPAT into the dorsal and median raphe nuclei on core temperature in the rat. *Psychopharmacology* 103, 291–296
93. Heisler, L.K. *et al.* (1998) Elevated anxiety and antidepressant-like responses in serotonin 5-HT<sub>1A</sub> receptor mutant mice. *Proc. Natl. Acad. Sci. U. S. A.* 95, 15049–15054
94. Richardson-Jones, J.W. *et al.* (2010) 5-HT<sub>1A</sub> autoreceptor levels determine vulnerability to stress and response to antidepressants. *Neuron* 65, 40–52
95. Hodges, M.R. *et al.* (2008) Defects in breathing and thermoregulation in mice with near-complete absence of central serotonin neurons. *J. Neurosci.* 28, 2495–2505
96. McGlashan, J.M. *et al.* (2015) Central serotonergic neurons activate and recruit thermogenic brown and beige fat and regulate glucose and lipid homeostasis. *Cell Metab.* 21, 692–705
97. Ray, R.S. *et al.* (2011) Impaired respiratory and body temperature control upon acute serotonergic neuron inhibition. *Science* 333, 637–642
98. Alenina, N. *et al.* (2009) Growth retardation and altered autonomic control in mice lacking brain serotonin. *Proc. Natl. Acad. Sci. U. S. A.* 106, 10332–10337
99. Cerpa, V. *et al.* (2014) Diphtheria toxin treatment of Pet-1-Cre floxed diphtheria toxin receptor mice disrupts thermoregulation without affecting respiratory chemoreception. *Neuroscience* 279, 65–76
100. Karasov, W.H. (1992) Daily energy expenditure and the cost of activity in mammals. *Integr. Comp. Biol.* 32, 238–248
101. van der Kooy, D. and Hattori, T. (1980) Dorsal raphe cells with collateral projections to the caudate-putamen and substantia nigra: a fluorescent retrograde double labeling study in the rat. *Brain Res.* 186, 1–7
102. Kazakov, V.N. *et al.* (1992) Sources of cortical, hypothalamic and spinal serotonergic projections: topical organization in the dorsal raphe nucleus. *Neurophysiology* 24, 69–76
103. Hillegaart, V. (1990) Effects of local application of 5-HT and 8-OH-DPAT into the dorsal and median raphe nuclei on motor activity in the rat. *Physiol. Behav.* 48, 143–148
104. Eagle, D.M. *et al.* (2009) Serotonin depletion impairs waiting but not stop-signal reaction time in rats: implications for theories of

- the role of 5-HT in behavioral inhibition. *Neuropsychopharmacology* 34, 1311–1321
105. Correia, P.A. *et al.* (2017) Transient inhibition and long-term facilitation of locomotion by phasic optogenetic activation of serotonin neurons. *Elife* 6, e20975
  106. Seo, C. *et al.* (2019) Intense threat switches dorsal raphe serotonin neurons to a paradoxical operational mode. *Science* 363, 538–542
  107. Ren, J. *et al.* (2018) Anatomically defined and functionally distinct dorsal raphe serotonin sub-systems. *Cell* 175, 472–487
  108. Marcinkiewicz, C.A. *et al.* (2016) Serotonin engages an anxiety and fear-promoting circuit in the extended amygdala. *Nature* 537, 97–101
  109. Fornal, C.A. *et al.* (1996) A subgroup of dorsal raphe serotonergic neurons in the cat is strongly activated during oral-buccal movements. *Brain Res.* 716, 123–133
  110. Waterhouse, B.D. *et al.* (2004) Sensorimotor-related discharge of simultaneously recorded, single neurons in the dorsal raphe nucleus of the awake, unrestrained rat. *Brain Res.* 1000, 183–191
  111. Kawashima, T. *et al.* (2016) The serotonergic system tracks the outcomes of actions to mediate short-term motor learning. *Cell* 167, 933–946
  112. Green, J.A. (2011) The heart rate method for estimating metabolic rate: review and recommendations. *Comp. Biochem. Physiol. A. Mol. Integr. Physiol.* 158, 287–304
  113. Robinson, S.E. *et al.* (1986) A GABA cardiovascular mechanism in the dorsal raphe of the rat. *Neuropharmacology* 25, 611–615
  114. Gradin, K. *et al.* (1992) Substance P injection into the dorsal raphe increases blood pressure and serotonin release in hippocampus of conscious rats. *Eur. J. Pharmacol.* 218, 363–367
  115. Bandler, R. and Shipley, M.T. (1994) Columnar organization in the midbrain periaqueductal gray: modules for emotional expression? *Trends Neurosci.* 17, 379–389
  116. Hahn, T.M. *et al.* (1998) Coexpression of AgRP and NPY in fasting-activated hypothalamic neurons. *Nat. Neurosci.* 1, 271–272
  117. Balthasar, N. *et al.* (2004) Leptin receptor signaling in POMC neurons is required for normal body weight homeostasis. *Neuron* 42, 983–991
  118. Kristensen, P. *et al.* (1998) Hypothalamic CART is a new anorectic peptide regulated by leptin. *Nature* 393, 72–76
  119. Wang, Q. *et al.* (2013) Arcuate AgRP neurons mediate orexigenic and glucoregulatory actions of ghrelin. *Mol. Metab.* 3, 64–72
  120. Krashes, M.J. *et al.* (2014) An excitatory paraventricular nucleus to AgRP neuron circuit that drives hunger. *Nature* 507, 238–242
  121. Watabe-Uchida, M. *et al.* (2012) Whole-brain mapping of direct inputs to midbrain dopamine neurons. *Neuron* 74, 858–873
  122. Beier, K.T. *et al.* (2015) Circuit architecture of VTA dopamine neurons revealed by systematic input-output mapping. *Cell* 162, 622–634
  123. Faget, L. *et al.* (2016) Afferent inputs to neurotransmitter-defined cell types in the ventral tegmental area. *Cell Rep.* 15, 2796–2808
  124. de Jong, J.W. *et al.* (2019) A neural circuit mechanism for encoding aversive stimuli in the mesolimbic dopamine system. *Neuron* 101, 133–151
  125. Rompre, P.P. and Miliaressis, E. (1985) Pontine and mesencephalic substrates of self-stimulation. *Brain Res.* 359, 246–259
  126. Fletcher, P.J. *et al.* (1993) Conditioned place preference induced by microinjection of 8-OH-DPAT into the dorsal or median raphe nucleus. *Psychopharmacology* 113, 31–36
  127. Shin, R. and Ikemoto, S. (2010) The GABA<sub>B</sub> receptor agonist baclofen administered into the median and dorsal raphe nuclei is rewarding as shown by intracranial self-administration and conditioned place preference in rats. *Psychopharmacology (Berl)* 208, 545–554
  128. Land, B.B. *et al.* (2009) Activation of the kappa opioid receptor in the dorsal raphe nucleus mediates the aversive effects of stress and reinstates drug seeking. *Proc. Natl. Acad. Sci. U. S. A.* 106, 19168–19173
  129. Zhong, W. *et al.* (2017) Learning and stress shape the reward response patterns of serotonin neurons. *J. Neurosci.* 37, 8863–8875
  130. Li, Y. *et al.* (2016) Serotonin neurons in the dorsal raphe nucleus encode reward signals. *Nat. Commun.* 7, 10503
  131. Miyazaki, K. *et al.* (2011) Activation of dorsal raphe serotonin neurons underlies waiting for delayed rewards. *J. Neurosci.* 31, 469–479
  132. McDevitt, R.A. *et al.* (2014) Serotonergic versus non-serotonergic dorsal raphe projection neurons: differential participation in reward circuitry. *Cell Rep.* 8, 1857–1869
  133. Fonseca, M.S. *et al.* (2015) Activation of dorsal raphe serotonergic neurons promotes waiting but is not reinforcing. *Curr. Biol.* 25, 306–315
  134. Miyazaki, K.W. *et al.* (2014) Optogenetic activation of dorsal raphe serotonin neurons enhances patience for future rewards. *Curr. Biol.* 24, 2033–2040
  135. Schweimer, J.V. and Ungless, M.A. (2010) Phasic responses in dorsal raphe serotonin neurons to noxious stimuli. *Neuroscience* 171, 1209–1215
  136. Cohen, J.Y. *et al.* (2015) Serotonergic neurons signal reward and punishment on multiple timescales. *Elife* 4, e06346
  137. Wang, H.L. *et al.* (2019) Dorsal raphe dual serotonin-glutamate neurons drive reward by establishing excitatory synapses on VTA mesoaccumbens dopamine neurons. *Cell Rep.* 26, 1128–1142
  138. Qi, J. *et al.* (2014) A glutamatergic reward input from the dorsal raphe to ventral tegmental area dopamine neurons. *Nat. Commun.* 5, 5390
  139. Lin, R. *et al.* (2020) The raphe dopamine system controls the expression of incentive memory. *Neuron* 106, 498–514
  140. Chalmers, D.T. *et al.* (1995) Localization of novel corticotropin-releasing factor receptor (CRF2) mRNA expression to specific subcortical nuclei in rat brain: comparison with CRF1 receptor mRNA expression. *J. Neurosci.* 15, 6340–6350
  141. Pernar, L. *et al.* (2004) Selective activation of corticotropin-releasing factor-2 receptors on neurochemically identified neurons in the rat dorsal raphe nucleus reveals dual actions. *J. Neurosci.* 24, 1305–1311
  142. Liu, R.J. *et al.* (2002) Hypocretins (orexins) regulate serotonin neurons in the dorsal raphe nucleus by excitatory direct and inhibitory indirect actions. *J. Neurosci.* 22, 9453–9464
  143. Marcus, J.N. *et al.* (2001) Differential expression of orexin receptors 1 and 2 in the rat brain. *J. Comp. Neurol.* 435, 6–25
  144. Jolas, T. and Aghajanian, G.K. (1996) Neurotensin excitation of serotonergic neurons in the dorsal raphe nucleus of the rat in vitro. *Eur. J. Neurosci.* 8, 153–161
  145. Moyses, E. *et al.* (1987) Distribution of neurotensin binding sites in rat brain: a light microscopic radioautographic study using monoiodo [<sup>125</sup>I]Tyr<sup>3</sup>-neurotensin. *Neuroscience* 22, 525–536
  146. Xu, Z.Q. *et al.* (1998) Galanin-5-hydroxytryptamine interactions: electrophysiological, immunohistochemical and in situ hybridization studies on rat dorsal raphe neurons with a note on galanin R1 and R2 receptors. *Neuroscience* 87, 79–94
  147. Lu, X. *et al.* (2005) A role for galanin in antidepressant actions with a focus on the dorsal raphe nucleus. *Proc. Natl. Acad. Sci. U. S. A.* 102, 874–879
  148. Lacoste, B. *et al.* (2006) Immunocytochemical evidence for the existence of substance P receptor (NK1) in serotonin neurons of rat and mouse dorsal raphe nucleus. *Eur. J. Neurosci.* 23, 2947–2958
  149. Sergeev, V. *et al.* (1999) Serotonin and substance P co-exist in dorsal raphe neurons of the human brain. *Neuroreport* 10, 3967–3970
  150. Van den Bergh, P. *et al.* (1988) Neurons containing a N-terminal sequence of the TRH-prohormone (preproTRH53-74) are present in a unique location of the midbrain periaqueductal gray of the rat. *Brain Res.* 461, 53–63
  151. Dougalis, A.G. *et al.* (2012) Functional properties of dopamine neurons and co-expression of vasoactive intestinal polypeptide in the dorsal raphe nucleus and ventro-lateral periaqueductal grey. *Eur. J. Neurosci.* 36, 3322–3332
  152. Moss, M.S. *et al.* (1983) The peptidergic organization of the cat periaqueductal gray. I. The distribution of immunoreactive

- enkephalin-containing neurons and terminals. *J. Neurosci.* 3, 603–616
153. Hurd, Y.L. and Fagergren, P. (2000) Human cocaine-and-amphetamine-regulated transcript (CART) mRNA is highly expressed in limbic- and sensory-related brain regions. *J. Comp. Neurol.* 425, 583–598
154. de Quidt, M.E. and Emson, P.C. (1986) Distribution of neuropeptide Y-like immunoreactivity in the rat central nervous system-II. Immunohistochemical analysis. *Neuroscience* 18, 545–618
155. Segerson, T.P. *et al.* (1987) Localization of thyrotropin-releasing hormone prohormone messenger ribonucleic acid in rat brain in situ hybridization. *Endocrinology* 121, 98–107